

**QUALITATIVE ANALYSIS AND ANTIOXIDANT POTENTIAL OF VANDA
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ABSTRACT

Vanda roxburghii R.Br. is a potent medicinal epiphytic orchid used in the Indian system of medicine. Benzene extract of *Vanda roxburghii* R.Br. leaf showed the presence of terpenoid, flavonoid, alkaloid, saponin, tannin and steroid; ethanol extract showed the presence of terpenoid, flavonoid, alkaloid, tannin and steroid; acetone extract showed the presence of terpenoid, alkaloid, tannin, steroid and chloroform extract showed the presence of flavonoid, reducing sugar, alkaloid and steroid. DPPH radical scavenging activity of *Vanda roxburghii* R.Br. leaf extracts varied from 17.98 % \pm 0.570 (20 μ l) of chloroform extract to 61.91% \pm 1.196 (100 μ l) of ethanol extract and hydroxyl radical scavenging activity of *Vanda roxburghii* R.Br. leaf extracts varied from 11.98 % \pm 0.789 (100 μ l) of chloroform extract to 58.5 % \pm 0.475 (500 μ l) of ethanol extract highlight the medicinal importance of the sampling plant.

KEYWORDS: Orchid, *Vanda roxburghii*, Phytochemicals, antioxidant.**INTRODUCTION**

Orchidaceae is a diverse and widespread family of flowering plants with blooms that are often colourful and often fragrant commonly known as the orchid family (White and Sharma, 2000). India is one of the richest orchid habitats about 2,500 species in 167 genera represented in six sub-families, 17 tribes and 30 sub-tribes (Hedge, 1997). Phytochemicals are biologically active, naturally occurring chemical compounds found in plants, which provide health benefits for humans than those attributed to macronutrients and micronutrients (Hasler *et al.*, 1999). Roots of *V. roxburghii* were reported to possess antibacterial, antitubercular properties, anti-inflammatory activity and aphrodisiac activity (Das *et al.*, 1967; Lawler, 1984). Antioxidants are believed to play a very important role in the body defense system against reactive oxygen species (ROS), which are the harmful by products generated during normal cell aerobic respiration (Rao *et al.*, 2012).

MATERIALS AND METHODS**Collection of Plant Material**

The epiphytic orchid *Vanda roxburghii* R. Br. was collected from Agastheeswaram Taluk at an altitude of about 250 feet of Kanyakumari District, the southernmost end of the peninsular India lies between 8°-20° north of the equator and between 70°-85° in

longitude. Photographs of the vegetative and reproductive (inflorescence) parts were compared with the description published in orchids of Nilgiris (Joseph, 1987).

Processing and Preparation of extracts

The freshly collected *Vanda roxburghii* R.Br. leaf was harvested and properly washed in tap water and then rinsed in sterile distilled water. The harvested leaves were dried in the hot air oven at 40° C for 3 days and the dried leaves were pulverized using sterile laboratory mortar and pestle to obtain a powdered form. The powdered samples were stored in airtight glass containers for further analysis. The dried powder of *Vanda roxburghii* R. Br. leaf was extracted with benzene, ethanol, acetone and chloroform in Soxhlet extractor for 72 hours and after exhaustive extraction, the leaf extracts were filtered with the help of rotary evaporator.

METHODOLOGY

Phytochemical constituents of *Vanda roxburghii* R.Br. leaf extracts was determined as per the standard procedures (Harborne, 1973). Antioxidant Activity was determined as per the standard procedure (DPPH radical scavenging activity of Yohozowa *et al.*, 1998 and

hydroxyl radical scavenging activity of Elizabeth *et al.*, 1990).

RESULTS AND DISCUSSION

Green plants synthesis and preserve variety of biochemical products many of which are extractable and are used as chemical feed stocks or as raw material for various scientific investigations. Suja and Williams, (2016) reported that the ethanol extract of *Acampe praemorsa* (Roxb) leaves revealed the presence of flavonoid, phenol, tannin and steroid constituents. The result of phytochemical activities showed that bioactive compounds such as alkaloid, terpenoids, flavonoids, phenols, tannins, steroids and glycosides were present in the whole plant extracts of *V. tessellata* (Bakul Bhattacharjee *et al.*, 2014). Phytochemical analysis of *Vanda tessellata* (Roxb.) Hook. showed the presence of flavonoid, glycoside, terpenoid and tannin (Maridass *et al.*, 2008). The present study revealed that the benzene extract of *Vanda roxburghii* R.Br. leaf showed the presence of terpenoid, flavonoid, alkaloid, saponin, tannin and steroid; ethanol extract showed the presence of terpenoid, flavonoid, alkaloid, tannin and steroid; acetone extract showed the presence of terpenoid, alkaloid, tannin, steroid and chloroform extract showed the presence of flavonoid, reducing sugar, alkaloid and steroid.

An antioxidant is a chemical compound that inhibits the oxidation of other molecules. 1, 1-diphenyl-2-picryl hydroxyl is a stable free radical scavenging activity react with DPPH reduce it to DPPH-H and as consequence, the absorbance decreases. DPPH radical scavenging activity of *Cottonia peduncularis* of acetone leaf extract showed highest percentage of inhibition (53.18%) (Nagananda *et al.*, 2013). DPPH radical scavenging of the benzene extract of *Vanda roxburghii* R.Br. leaf varied from $25.29\% \pm 1.618$ (20 μ l) to $59.29\% \pm 1.255$ (100 μ l); ethanol extract varied from $27.28\% \pm 0.196$ (25 μ l) to $61.91\% \pm 1.196$ (100 μ l); acetone extract varied from $20.98 \pm 1.618\%$ (20 μ l) to $50.86\% \pm 0.287$ (100 μ l); chloroform extract varied from $17.98\% \pm 0.570$ (20 μ l) to $45.87\% \pm 0.287$ (100 μ l) and antioxidant potential of the standard antioxidant ascorbic acid varied from $28.17\% \pm 0.672$ (20 μ l) to $67.17\% \pm 0.490$ (100 μ l) (Fig: 1).

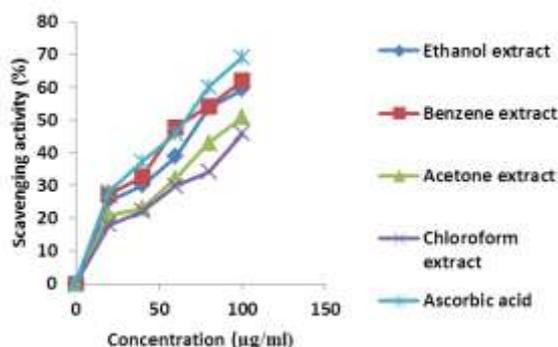


Fig. 1: DPPH Radical Scavenging.

Hydroxyl radical scavenging is an extremely reactive free radical formed in biological system. Uddin *et al.*, (2015) reported that the *Vanda roxburghii* chloroform extract posses potential hydroxyl radical scavenging activity. Ethanol extract of *Vanda roxburghii* R.Br. leaf varied from $18.12\% \pm 0.461$ (100 μ l) to $58.5\% \pm 0.475$ (500 μ l); benzene extract varied from $13.45\% \pm 0.280$ (100 μ l) to $43.65\% \pm 0.667$ (500 μ l); acetone extract varied from $12.8\% \pm 0.879$ (100 μ l) to $40.78\% \pm 0.787$ (500 μ l); chloroform extract of *Vanda roxburghii* R.Br. leaf varied from $11.98\% \pm 0.789$ (100 μ l) to $39.76\% \pm 0.678$ (500 μ l) and antioxidant potential of the standard antioxidant ascorbic acid varied from $19.98\% \pm 0.903$ (100 μ l) to $63.82\% \pm 0.000$ (500 μ l) (Fig: 2).

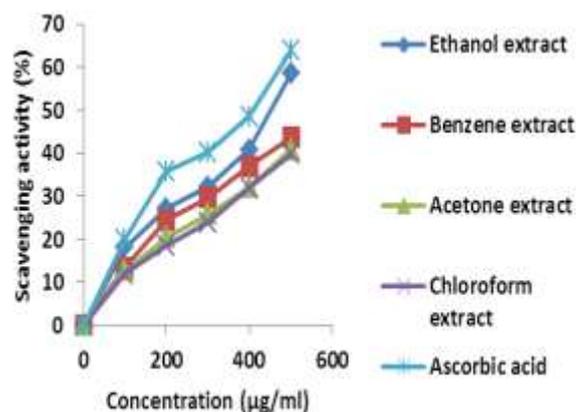


Fig. 2: Hydroxyl Radical Scavenging.

CONCLUSION

The phytochemical constituents and antioxidant potential of the sampling plants justified the traditional use of the plants further experiments are required to elucidate their mechanism of action at cellular and molecular levels.

REFERENCES

1. Bakul Bhattacharjee, Touhidul Islam, Zamilur Rahman and Shahinul Islam. 2014. Antimicrobial activity and phytochemical screening of whole plant extracts of *Vanda Tessellata* (Roxb.). *World Journal of Pharmacy and Pharmaceutical Sciences*, 4(1): 72-83.
2. Das, S, Bhattacharya, A. and A.K. Bhattacharya. 1967. Active constituents of *Vanda roxburghii* R. Br. *J. Indian Chem. Soc.*, 44: 804-5.
3. Elizabeth, K. and Rao, M.N.A. 1990. Oxygen radical scavenging activity of *Curcumin*. *Int J. Pharm*, 58: 237-240.
4. Harborne J.B. 1973. *Phytochemical methods: A guide to modern techniques of plant analysis*, 13th Ed. *Chapman and Hall, Ltd. London*, 5-15.
5. Hasler, C.M. and Blumberg, J.B. 1999. Symposium on Phytochemicals, Biochemistry and Physiology. *Journal of Nutrition*, 129: 756-757.
6. Hedge, S.N. 1997. *Orchid Wealth of India. Proceedings of Indian National Science Academy*, 63: 229-244.

7. Joseph. J. 1987. Orchids of Nilgiris, Printed by the Director Botanical Survey of India, New Delhi, India.
8. Lawler, L.J. 1984. Ethnobotany of the *Orchidaceae*. In: Arditti J(ed) *Orchid Biology: Review and Perspectives-3*. Cornell University Press, Ithaca, 27- 149.
9. Maridass, M., Zahir Hussain, B and Raju, G. 2008. Phytochemical Survey of Orchids in the Tirunelveli Hills of South India. *Ethnobotanical Leaflets*, 12: 705-12.
10. Mary Suja, R. and Christudhas Williams, B. 2016. Micropropagation, Phytochemical Screening and Antioxidant Potential of a Wild Epiphytic Orchid *Acampe praemorsa* (Roxb) of Kanyakumari District, India". *European Journal of Pharmaceutical and Medical Research*, 572-576.
11. Nagananda, G.S. and N. Satishchandra. 2013. Antimicrobial activity of cold and hot successive Pseudobulb extract of *Flickingeria nodosa* (Dalz.) seidenf. *Pak. J. Bio. Sci*, 16(20): 1189-1193.
12. Rao, S.G., Kiran, G., Ramakrishna, P.G., Marella, P., Aviv, R.B.Y. and C. Swetha. 2012. Evaluation of In vivo antioxidant activity of *vanda tessellata* Roxb. in albino wistar rats. *Int. J. Exp. Pharm*, 2(2): 71-74.
13. Uddin, N.M.D., Afrin, R.M.D., Uddin, J.M.D., Uddin, J., Alam, A. H. M. K., Rahman, A.A and G. Sadik. 2015. *Vanda roxburghii* chloroform extract as potential source of polyphenols with antioxidant and cholinesterase inhibitory activities: identification of a strong phenolic antioxidant. *BMC Complementary and Alternative Medicine*, 15(195): 3-9.
14. White, K.J. and B. Sharma 2000. Wild orchids in Nepal the guide to the Himalayan orchids of the Tribhuvan Rajpath and Chitwan Jungle. Bangkok. Thailand: *white lotus press*.